

## Synthesis of 6-Chloro-16-methylene-17 $\alpha$ -hydroxy-21-fluoro-4,6-pregnadiene-3,20-dione 17-Acetate, a Potent Progestational Agent

T. L. POPPER,\* F. E. CARLON, E. L. SHAPIRO, AND R. NERI

*Natural Products Research Department, and Physiology and Biochemistry Department,  
Schering Corporation, Bloomfield, New Jersey 07003*

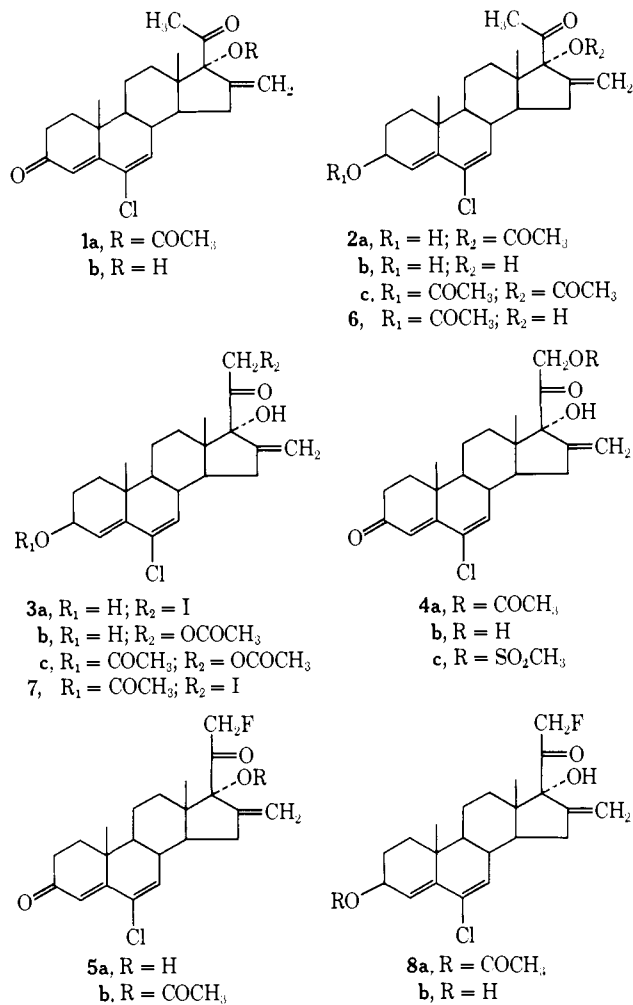
Received March 24, 1970

The synthesis of the title compound **5b**, a potent progestational agent, is reported. The intermediates **2a** and **2c** also possessed high progestational activities when tested intramuscularly or orally in the rabbit.

The progestational potentiating effect of the 21-fluoro group in 17 $\alpha$ -acetoxyprogesterone derivatives has been described.<sup>1</sup> It was reported recently that 6-chloro-16-methylene-17 $\alpha$ -hydroxy-4,6-pregnadiene-3,20-dione 17-acetate (**1a**) has approximately twice the progestational activity of the corresponding 16-unsubstituted compound.<sup>2</sup> We now report the synthesis of 6-chloro-16-methylene-17 $\alpha$ -hydroxy-21-fluoro-4,6-pregnadiene-3,20-dione 17-acetate (**5b**) in which both of the structural features known to increase progestational activity are combined in the same molecule.

A straightforward sequence for the preparation of the desired 21-fluoro compounds **5a** and **5b** was the utilization of the 21-bromo and/or-iodo derivatives of **1b** or **1a** for conversion into the corresponding 21-fluoro steroid by known methods. However, all efforts to brominate or iodinate **1b** or its 1-dehydro derivative<sup>3</sup> selectively at C<sub>21</sub> failed, halogenation at C<sub>2</sub> always preceded halogenation at C<sub>21</sub>.

A second approach to the desired **5b** involved protection of the 3-ketone by reduction, to avoid halogenation of ring A. This method seemed to be quite attractive since the mesylate **4c** would be an equally suitable precursor for the 21-chloro and 21-bromo analogs of **5b**, also of biological interest. Reduction of **1a** with NaBH<sub>4</sub> in cold CH<sub>2</sub>Cl<sub>2</sub>-MeOH gave in high yield the 3 $\beta$ -hydroxy **2a**, which was hydrolyzed with KOH in MeOH-H<sub>2</sub>O affording the diol **2b**. Brief exposure of **2b** to I<sub>2</sub>, CaO, and azobis-2-methylpropionitrile<sup>4</sup> gave the impure 21-iodo compound **3a**. Short reaction time in the 21-iodination step was essential to minimize the unwanted oxidation by I<sub>2</sub> of the 3 $\beta$ -hydroxy- $\Delta^{4,6}$ -system. Even after brief exposure, however, some oxidation did occur, and the resulting  $\Delta^{4,6}$ -3-one rapidly underwent iodination at C<sub>2</sub> followed by elimination of HI in the basic medium, resulting in the formation of the undesired  $\Delta^{1,4,6}$ -3-one. The reaction mixture, without purification, was treated with Et<sub>3</sub>N in refluxing AcOH in order to convert the 21-iodo **3a** into the 21-acetoxy **3b**. The crude reaction product was treated with activated MnO<sub>2</sub> in CHCl<sub>3</sub> to regenerate the 4,6-dienone system. Chromatogra-



phy of the MnO<sub>2</sub> oxidation product over Florisil afforded crystalline **4a** in about 25% yield from **2b**.

Hydrolysis of **4a** with KOH in MeOH-H<sub>2</sub>O resulted in the diol **4b**, which on treatment with MsCl in pyridine at 0° gave the chromatographically homogeneous mesylate **4c**. When **4c** was treated with anhydrous KF in DMSO at 110°,<sup>5</sup> none of the desired **5a** could be isolated from the reaction mixture, under these conditions only decomposition of **4c** occurred, while milder conditions resulted in the recovery of starting material.

In order to overcome the difficulties in the replacement of the mesylate with KF, the reaction sequence

\* To whom correspondence should be addressed.

(1) (a) R. L. Elton, *Proc. Soc. Exptl. Biol. Med.*, **101**, 677 (1959); (b) V. A. Drill, *Fed. Proc., Fed. Amer. Soc. Exptl. Biol.*, **18**, 1040 (1959); (c) C. G. Bergstrom, P. B. Sollman, R. T. Nicholson, and R. M. Dodson, *J. Amer. Chem. Soc.*, **82**, 2322 (1960).

(2) Z. Cekan, M. Seda, J. Mikulaskova, and K. Syhora, *Steroids*, **8**, 205 (1966).

(3) E. L. Shapiro, T. L. Popper, L. Weber, R. Neri, and H. L. Herzog, *J. Med. Chem.*, **12**, 631 (1969).

(4) E. S. Rothmann, T. Perlstein, and M. E. Wall, *J. Org. Chem.*, **25**, 1966 (1960), and ref cited therein.

(5) J. E. Herz, J. Fried, P. Grabovich, and E. F. Sabo, *J. Amer. Chem. Soc.*, **78**, 4812 (1956), reported these conditions for the conversion of 21-mesylate derivatives of corticoids into the corresponding 21-fluoro-21-desoxycorticoids in the 16-unsubstituted series.

was somewhat modified. Acetylation of **2b** with  $\text{Ac}_2\text{O}$  in pyridine in order to minimize the oxidation of the  $\beta$ -hydroxy group during the iodination step gave the 3-monoacetate **6**. Brief treatment of **6** with 2 equiv of  $\text{I}_2$  in the presence of  $\text{CaO}$  and azobis-2-methylpropionitrile gave a mixture of products of which the 21-iodo compounds **7** and **3a** predominated. This mixture, without purification was treated with 50% aq  $\text{AgF}$  in  $\text{MeCN}$  at reflux<sup>6</sup> and the resulting product, predominantly a mixture of **8a** and **8b**, was treated with  $\text{KOH}$  in  $\text{MeOH-H}_2\text{O}$  and after chromatography over neutral alumina the desired **8b** was obtained in an overall yield of 20–25% from **6**. Dehydrogenation of **8b** with activated  $\text{MnO}_2$  in  $\text{DMF}$  afforded **5a** which was acetylated with  $\text{AcOH}$ , trifluoroacetic anhydride (TFAA) and *p*-TSA· $\text{H}_2\text{O}$ <sup>7</sup> affording the desired **5b**.

Since 6-chloro- $\beta$ ,17 $\alpha$ -dihydroxy-4,6-pregnadiene-20-one diacetate was reported to be a potent progestational agent,<sup>8</sup> the 16-methylene analog **2c** was prepared by acetylation of **2a** with  $\text{Ac}_2\text{O}$  in pyridine.

**Biological Activity.**—Table I lists the intramuscular (im) and oral progestational activities of **1a**, **2a**, **2c** and **5b**. As expected, introduction of the 21-fluoro substituent increased the im progestational activity of **1a** about 1.4-fold.

TABLE I  
PROGESTATIONAL ACTIVITY<sup>a</sup>

Compd	Route of administration <sup>b</sup>	
	Im	Oral
9 <sup>c</sup>		2.8
1a	77	55
2a	69.8	13.7
2c	34.5	47
5b	105.4	32.5

<sup>a</sup> Progesterone (im) = 1. <sup>b</sup> Progestational activity was determined in immature rabbits by the method of M. K. McPhail, *J. Physiol. (London)*, **83**, 145 (1934). The compounds were dissolved in sesame oil for im administration or suspended in an aq medium (0.9%  $\text{NaCl}$ , 0.5%  $\text{CM-cellulose}$ , 0.4% polysorbate 80, and 0.9%  $\text{PhCH}_2\text{OH}$ ) for oral administration. Progesterone in sesame oil was always given im. The statistical analysis for the progestational assays utilized the randomized Bloch analysis of variance with Dunnett's and Duncan's multiple comparison procedure (see G. Miller, Jr., "Simultaneous Statistical Interference," McGraw-Hill Book Co., Inc., New York, N. Y., 1967). <sup>c</sup> 17 $\alpha$ -Ethinyl-19-nortestosterone.

## Experimental Section<sup>9</sup>

**6-Chloro-16-methylene- $\beta$ ,17 $\alpha$ -dihydroxy-4,6-pregnadiene-20-one 17-Acetate (2a).**—A soln of 6-chloro-16-methylene-17 $\alpha$ -hydroxy-4,6-pregnadiene-3,20-dione 17-acetate (**1a**) (8.6 g) in  $\text{MeOH}$  (40 ml) and  $\text{CH}_2\text{Cl}_2$  (40 ml) was stirred with  $\text{NaBH}_4$  (3.5 g) and  $\text{H}_2\text{O}$  (10 ml) at 0° under  $\text{N}_2$  for 20 min. The solution was neutralized with dil  $\text{AcOH}$  and extracted with  $\text{CH}_2\text{Cl}_2$ . After drying it was evaporated to a residue. The residue was dissolved in 500 ml of  $\text{MeOH}$  and a 100-ml aliquot of this soln was concd to low vol when crystn occurred yielding 1.10 g of

**2a**: mp 220–223°;  $[\alpha]_D - 215^\circ$ ;  $\lambda_{\text{max}}$  236  $\text{m}\mu$  ( $\epsilon$  20,900), 243 (24,200), 251 (16,300);  $\delta$  4.30 ( $\text{C}_{32}\text{-H}$ , m, 20 Hz wide) ppm. *Anal.* ( $\text{C}_{22}\text{H}_{31}\text{O}_3\text{Cl}$ ) C, H, Cl.

**6-Chloro-16-methylene- $\beta$ ,17 $\alpha$ -dihydroxy-4,6-pregnadiene-20-one (2b).**—A soln of **2a** (500 mg) in  $\text{MeOH}$  (30 ml) was stirred with 1 *N*  $\text{KOH}$  (5 ml) under  $\text{N}_2$  for 20 hr. The soln was neutralized with  $\text{AcOH}$  and dild with  $\text{H}_2\text{O}$  and the ppt collected by filtration. Crystn from  $\text{Me}_2\text{CO-i-Pr}_2\text{O}$  gave 242 mg of **2b**: mp 163–165°;  $[\alpha]_D - 130^\circ$ ;  $\lambda_{\text{max}}$  235  $\text{m}\mu$  ( $\epsilon$  20,320), 241 (22,700), 250 (15,750). *Anal.* ( $\text{C}_{22}\text{H}_{29}\text{O}_3\text{Cl}$ ) C, H, Cl.

**6-Chloro-16-methylene- $\beta$ ,17 $\alpha$ -dihydroxy-4,6-pregnadiene-20-one 3-Acetate (6).**—A soln of **2b** (1.15 g) in  $\text{C}_6\text{H}_5\text{N}$  (4 ml) was allowed to stand with  $\text{Ac}_2\text{O}$  (4 ml) at 20° for 3 hr, followed then for 10 min on the steam bath. The reaction mixture was added to ice- $\text{H}_2\text{O}$ , the ppt collected and dried. Crystn from  $\text{MeOH-i-Pr}_2\text{O-C}_6\text{H}_{14}$  yielded 779 mg of **6**: mp 162–163°;  $[\alpha]_D - 136^\circ$ ;  $\lambda_{\text{max}}$  235  $\text{m}\mu$  ( $\epsilon$  20,400), 240 (22,050), 249 (15,350). *Anal.* ( $\text{C}_{24}\text{H}_{31}\text{O}_5\text{Cl}$ ) C, H, Cl.

**6-Chloro-16-methylene- $\beta$ ,17 $\alpha$ -dihydroxy-4,6-pregnadiene-20-one 3,17-Diacetate (2c).**—A  $\text{C}_6\text{H}_5\text{N}$  soln (2 ml) of the crude product of **2a** (1 g) was allowed to stand with  $\text{Ac}_2\text{O}$  (3 ml) for 60 hr. The reaction mixture was added to ice- $\text{H}_2\text{O}$ , the ppt collected and dried. Crystallization from  $\text{Et}_2\text{O-C}_6\text{H}_{14}$  gave 767 mg of **2c**: mp 194–196° dec;  $[\alpha]_D - 215^\circ$ ;  $\lambda_{\text{max}}$  236  $\text{m}\mu$  ( $\epsilon$  21,300), 243 (24,000), 251 (16,150). *Anal.* ( $\text{C}_{26}\text{H}_{33}\text{O}_5\text{Cl}$ ) C, H, Cl.

**6-Chloro-16-methylene-17 $\alpha$ ,21-dihydroxy-4,6-pregnadiene-3,20-dione 21-Acetate (4a).**—To a soln of **2b** (7.2 g, 19 mmoles) in  $\text{THF}$  (75 ml, passed through act. I. Alumina) and  $\text{MeOH}$  (35 ml) under  $\text{N}_2$  was added freshly fused  $\text{CaO}$  (12 g), azobis-2-methylpropionitrile (750 mg) and  $\text{I}_2$  (8.52 g, 33.5 mmoles). The reaction mixture was stirred for 7 min. The solids were removed by filtration, the filtrate added to  $\text{H}_2\text{O}$  containing  $\text{Na}_2\text{S}_2\text{O}_3$ . The ppt was collected, dried, and dissolved in  $\text{Me}_2\text{CO}$  (120 ml). A 100-ml part of this soln was heated at reflux with  $\text{Et}_3\text{N}$  (60 ml) and  $\text{AcOH}$  (40 ml) for 1 hr. The reaction mixture was cooled and added to ice- $\text{H}_2\text{O}$ . The ppt was collected, dried, dissolved in  $\text{CHCl}_3$  (500 ml), and stirred with activated  $\text{MnO}_2$  (15 g) for 1.5 hr. The solids were removed by filtration and the filtrate concentrated *in vacuo* to a residue which was chromatographed over Florisil (27 × 4 cm). Elution with  $\text{C}_6\text{H}_6\text{-CH}_2\text{Cl}_2$  (1:1) yielded, after crystn from  $\text{Et}_2\text{O}$ , 1.71 g of **4a**: mp 229–234° dec;  $[\alpha]_D + 6^\circ$ ;  $\lambda_{\text{max}}$  284  $\text{m}\mu$  ( $\epsilon$  20,500); nmr,  $\delta$  0.79 ( $\text{C}_{13}\text{-CH}_3$ ), 1.14 ( $\text{C}_{10}\text{-CH}_3$ ), 2.15 ( $\text{C}_{21}\text{-OCOCH}_3$ ), 4.92 and 5.10 ( $\text{C}_{20}\text{-CH}_2\text{O}$ ,  $J_{\text{gem}} = 18\text{ Hz}$ ), 5.13 and 5.28 ( $\text{C}_{16}\text{=CH}_2$ ), 6.27 ( $\text{C}_7\text{-H}$ ), and 6.30 ( $\text{C}_4\text{-H}$ ) ppm. *Anal.* ( $\text{C}_{24}\text{H}_{29}\text{O}_5\text{Cl}$ ) C, H, Cl.

**6-Chloro-16-methylene-17 $\alpha$ ,21-dihydroxy-4,6-pregnadiene-3,20-dione (4b).**—A soln of **4a** (1 g) in  $\text{MeOH}$  (10 ml) was allowed to stand with 1 *N* aq  $\text{KOH}$  (3 ml) under  $\text{N}_2$  at 20° for 2 hr. After neutralization with  $\text{AcOH}$  the soln was added to  $\text{H}_2\text{O}$ . The ppt was collected, dried, and chromatographed over silica gel (Baker, act. V, 56 × 2.5 cm). Elution with  $\text{CH}_2\text{Cl}_2\text{-Et}_2\text{O}$  (9:1) gave 373 mg of **4b** which was crystd from  $\text{MeOH-Et}_2\text{O}$ : mp 154–155°;  $[\alpha]_D - 9^\circ$ ;  $\lambda_{\text{max}}$  283  $\text{m}\mu$  ( $\epsilon$  21,400). *Anal.* ( $\text{C}_{22}\text{H}_{27}\text{O}_5\text{Cl}$ ) C, H, Cl.

**Attempted Conversion of 4b into 5a.**—A soln of **4b** (200 mg) in  $\text{C}_6\text{H}_5\text{N}$  (4 ml) and  $\text{MsCl}$  (0.3 ml) was allowed to stand at 0° for 2.5 hr. The soln was added to  $\text{H}_2\text{O}$ , the ppt collected and dried. The chromatographically homogeneous mesylate **4c** [ $\nu_{\text{max}}$  1738, 1665, 1355, 1170  $\text{cm}^{-1}$ ; nmr,  $\delta$  3.20 ( $\text{C}_{21}\text{-OSO}_2\text{CH}_3$ ), 5.11 and 5.39 ( $\text{C}_{20}\text{-CH}_2\text{O}$ ,  $J_{\text{gem}} = 18.5\text{ Hz}$ ), 5.14 and 5.27 ( $\text{C}_{16}\text{=CH}_2$ ) ppm] was added to  $\text{DMSO}$  (15 ml) containing  $\text{KF}$  (500 mg, freshly fused) and kept at 110° for 16 hr. The dark soln was cooled, dild with  $\text{H}_2\text{O}$ , and extd successively with  $\text{CH}_2\text{Cl}_2$  and  $\text{EtOAc}$ . The organic solns were combined, dried, and evaporated to a residue *in vacuo*. No **5a** could be detected, apparently complete decompn of **4c** occurred, as evidenced by tlc.

**6-Chloro-16-methylene- $\beta$ ,17 $\alpha$ -dihydroxy-21-fluoro-4,6-pregnadiene-20-one (8b).**—To a soln of **6** (11.168 g, 26.7 mmoles) in  $\text{THF}$  (120 ml, passed through act. I. Alumina) and  $\text{MeOH}$  (60 ml) under  $\text{N}_2$  was added freshly fused  $\text{CaO}$  (23 g), azobis-2-methylpropionitrile (700 mg) and  $\text{I}_2$  (13.6 g, 53.4 mmoles). The reaction mixture was stirred for 10 min. The solids were removed by filtration, the filtrate added to  $\text{H}_2\text{O}$  containing  $\text{Na}_2\text{S}_2\text{O}_3$  and extd with  $\text{CH}_2\text{Cl}_2$ . Evaporation of the solvent gave an oily residue which was dissolved in  $\text{MeCN}$  (250 ml) and heated at reflux with 50% aq  $\text{AgF}$  soln (18 ml) for 9 hr. The solids were removed by filtration, the filtrate concd to a residue *in vacuo* which was dissolved in  $\text{Me}_2\text{CO}$  (50 ml) and added to  $\text{H}_2\text{O}$ . The ppt was collected, washed, dissolved in  $\text{MeOH}$  (100 ml), and

(6) E. V. Jensen, R. J. Pratt, and P. Tannhauser, *J. Amer. Chem. Soc.*, **78**, 2658 (1956).

(7) E. L. Shapiro, L. Finckenor, H. Pluchet, L. Weber, C. H. Robinson, E. P. Oliveto, H. L. Herzog, I. I. A. Tabachnick, and E. Collins, *Steroids*, **9**, 143 (1967).

(8) C. Revesz, U. K. Banik, and F. Herr, *ibid.*, **10**, 291 (1967).

(9) Melting points are uncorrected. Rotations are in dioxane at 25° at about 1% concentration, uv spectra are of  $\text{MeOH}$  solutions, and ir spectra are in  $\text{Nujol}$  unless otherwise stated. The nmr spectra were measured on a Varian A 60-A spectrometer in  $\text{CDCl}_3$  ( $\text{Me}_4\text{Si}$ ). Solutions were dried over anhyd  $\text{Na}_2\text{SO}_4$ . Analyses were determined by the Physical Organic Department of the Schering Corp. Where analyses are indicated only by symbols of the elements, analytical results obtained for those elements were within  $\pm 0.4\%$  of the theoretical values.

irred with 1 *N* aq KOH (40 ml) under N<sub>2</sub> for 75 min. After neutralization with AcOH the soln was added to H<sub>2</sub>O. The ppt as collected, dried, and chromatographed over neutral alumina Woelm, act. V, 40 × 4 cm). Elution with CH<sub>2</sub>Cl<sub>2</sub> gave, after ystn from Me<sub>2</sub>CO-Et<sub>2</sub>O, 1.30 g of **8b**: mp 178–180°; [α]<sub>D</sub> 93°; λ<sub>max</sub> 237 mμ (ε 18,650), 244 (21,800), 252 (14,900); nr, δ 0.84 (C<sub>13</sub>-CH<sub>3</sub>), 1.09 (C<sub>10</sub>-CH<sub>3</sub>), 5.14 and 5.31 (C<sub>16</sub>=CH<sub>2</sub>), 13 and 5.35 (C<sub>20</sub>-CH<sub>2</sub>F, J<sub>HF</sub> = 48 Hz), 5.83 (C<sub>7</sub>-H), and 6.10 (C<sub>4</sub>-H) ppm. Anal. (C<sub>22</sub>H<sub>28</sub>O<sub>3</sub>ClF) C, H, Cl; F: calcd, 4.81; found, 4.39.

The mother liquor and the later fractions were rechromatographed over neutral alumina (Woelm, act. V, 20 × 4 cm). lution with C<sub>6</sub>H<sub>6</sub> gave an additional 793 mg of **8b**.

**6-Chloro-16-methylene-17α-hydroxy-21-fluoro-4,6-pregnadiene-3,20-dione 17-Acetate (5b).**—Trifluoroacetic anhydride (4 ml) was added to a soln of **5a** (607 mg) and *p*-TSA·H<sub>2</sub>O (60 mg) in AcOH (6 ml) dropwise at 10° in a period of 10 min under N<sub>2</sub>. The reaction mixture was allowed to warm up to 20° and stirred 5 hr. The soln was added to H<sub>2</sub>O, the ppt collected, washed, and dried. Crystn from CH<sub>2</sub>Cl<sub>2</sub>-*i*-Pr<sub>2</sub>O gave 384 mg of **5b**: mp 234–238° dec; [α]<sub>D</sub> -147°; λ<sub>max</sub> 284 mμ (ε 22,500); ν<sub>max</sub> 1747 (sh), 1670, 1607, 1245 cm<sup>-1</sup>; nmr, δ 0.82 (C<sub>13</sub>-CH<sub>3</sub>), 1.15 (C<sub>10</sub>-CH<sub>3</sub>), 2.20 (C<sub>17</sub>-OCOCH<sub>3</sub>), 4.98 and 5.06 (C<sub>20</sub>-CH<sub>2</sub>F, J<sub>HF</sub> = 47 Hz, J<sub>H<sub>2</sub>F</sub> = 47.5 Hz), 5.47 and 5.58 (C<sub>16</sub>=CH<sub>2</sub>), 6.26 (C<sub>7</sub>-H), and 6.30 (C<sub>4</sub>-H) ppm. Anal. (C<sub>24</sub>H<sub>28</sub>O<sub>4</sub>ClF) C, H, Cl, F.

mμ (ε 22,200). Anal. (C<sub>22</sub>H<sub>28</sub>O<sub>3</sub>ClF) C, H, Cl; F: calcd, 4.83; found, 4.39.

**6-Chloro-16-methylene-17α-hydroxy-21-fluoro-4,6-pregnadiene-3,20-dione 17-Acetate (5b).**—Trifluoroacetic anhydride (4 ml) was added to a soln of **5a** (607 mg) and *p*-TSA·H<sub>2</sub>O (60 mg) in AcOH (6 ml) dropwise at 10° in a period of 10 min under N<sub>2</sub>. The reaction mixture was allowed to warm up to 20° and stirred 5 hr. The soln was added to H<sub>2</sub>O, the ppt collected, washed, and dried. Crystn from CH<sub>2</sub>Cl<sub>2</sub>-*i*-Pr<sub>2</sub>O gave 384 mg of **5b**: mp 234–238° dec; [α]<sub>D</sub> -147°; λ<sub>max</sub> 284 mμ (ε 22,500); ν<sub>max</sub> 1747 (sh), 1670, 1607, 1245 cm<sup>-1</sup>; nmr, δ 0.82 (C<sub>13</sub>-CH<sub>3</sub>), 1.15 (C<sub>10</sub>-CH<sub>3</sub>), 2.20 (C<sub>17</sub>-OCOCH<sub>3</sub>), 4.98 and 5.06 (C<sub>20</sub>-CH<sub>2</sub>F, J<sub>HF</sub> = 47 Hz, J<sub>H<sub>2</sub>F</sub> = 47.5 Hz), 5.47 and 5.58 (C<sub>16</sub>=CH<sub>2</sub>), 6.26 (C<sub>7</sub>-H), and 6.30 (C<sub>4</sub>-H) ppm. Anal. (C<sub>24</sub>H<sub>28</sub>O<sub>4</sub>ClF) C, H, Cl, F.

**Acknowledgments.**—We are indebted to Dr. H. L. Herzog for helpful discussions, Mr. M. D. Yudis and Mr. J. Morton for interpretation of the nmr spectra.

## Glyoxalase Inhibitors. A Possible Approach to Anticancer Agents<sup>1</sup>

ROBERT VINCE\* AND SUSAN DALUGE<sup>2</sup>

Department of Medicinal Chemistry, College of Pharmacy,  
University of Minnesota, Minneapolis, Minnesota 55455

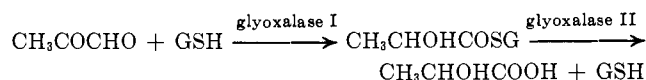
Received May 7, 1970

Some glyoxalase I enzyme inhibitors were assayed for cytotoxic activity against L1210 leukemia and KB cells in tissue culture. Sublethal concentrations of glyoxalase inhibitor caused a 14- to 18-fold increase in methylglyoxal toxicity in L1210 cells. A probable mechanism for the cytotoxic activity of methylglyoxal is discussed.

In a preliminary account of our work we presented our proposal that selective inhibition of the enzyme, glyoxalase I, may provide carcinostatic activity by preventing the metabolism of the cytotoxic ketoaldehyde, methylglyoxal, in tumor cells.<sup>3</sup> The carcinostatic activity of α-ketoaldehydes, including methylglyoxal, was first reported by French and Freedlander.<sup>4</sup> However, these agents are metabolized to the corresponding α-hydroxy acids by the glyoxalase system, thus obviating their use as effective anticancer agents. It has been well established that the cytotoxic methylglyoxal is the substrate for the glyoxalase enzymes and is converted to the nontoxic lactic acid in the presence of a cofactor, glutathione (GSH).<sup>5</sup> These facts, along with the observation by Stern<sup>6</sup> that the GSH concentration in cells rapidly increased just prior to cell division, suggests that the glyoxalase system may be involved in the regulation of cell growth by maintaining a proper concentration of methylglyoxal.<sup>3</sup> The high concentration of lactic acid<sup>7</sup> and the deficiency of methylglyoxal<sup>8</sup> in cancer cells further suggests that such cells, having lost the ability to maintain a proper balance of methylglyoxal, con-

tinue to grow at an uncontrolled rate. Recent interest in the possible role of methylglyoxal in cancer chemotherapy<sup>9</sup> has been concentrated on the preparation of ketoaldehydes. Since the toxicity of ketoaldehydes alone does not demonstrate the involvement of the glyoxalase system, we would like to report some results of our approach to this problem.

The glyoxalase system is widely distributed in cells of all forms of life<sup>10,11</sup> and comprises two enzymes which catalyze the reaction:



Since the inhibition of glyoxalase I may result in a buildup of methylglyoxal in cancer cells, our efforts have been concentrated on the preparation of inhibitors of this enzyme. Preliminary investigations demonstrated that S-alkyl derivatives of GSH cause potent competitive inhibition of glyoxalase I by taking advantage of a nonpolar region adjacent to the binding region of the enzyme.<sup>3</sup> A wide variety of S-alkyl and S-aryl glutathiones was prepared to investigate further the binding requirements of these inhibitors to the enzyme<sup>12</sup> and to provide for greater penetration of the cell membrane. The compounds were then investigated for cell kill of L1210 leukemia and KB cell cultures.<sup>13</sup> Since the S-alkyl derivatives seem to have difficulty in penetrating the cell membrane, only those S-aryl glutathiones

\* To whom correspondence should be addressed.

(1) This work was generously supported by Grant CA-10979 from the National Cancer Institute, U. S. Public Health Service, and a grant from the Minnesota Division of the American Cancer Society.

(2) Supported as a Postdoctoral trainee on U. S. Public Health Service Training Grant GM-01769 during the period 2-1-69 to 1-31-70.

(3) R. Vince and W. B. Wadd. *Biochem. Biophys. Res. Commun.*, **35**, 3 (1969).

(4) F. A. French and B. L. Freedlander, *Cancer Res.*, **18**, 172 (1958).

(5) E. Bricas and C. Fromageot, *Advan. Protein Chem.*, **8**, 1 (1953); E. Bricas in "Glutathione", S. P. Colowick, et al., Ed., Academic Press, New York, N. Y., 1954, p 165; W. E. Knox, *Enzymes*, **2**, 253 (1960).

(6) H. Stern, *Science*, **124**, 1292 (1956).

(7) B. Issekutz, "The Chemotherapy of Cancer," Akademiai Kiado, Budapest, 1969, pp 17–19.

(8) K. F. Lewis, E. H. Majane, and S. Weinhouse, *Cancer Res.*, **19**, 97 (1959).

(9) A. Szent-Györgyi, *Science*, **161**, 988 (1968).

(10) K. Lohman, *Biochem. Z.*, **254**, 332 (1932).

(11) F. G. Hopkins and E. J. Morgan, *Biochem. J.*, **194**, 119 (1952).

(12) R. Vince, S. Daluge, and W. B. Wadd, *J. Med. Chem.*, submitted for publication.

(13) We wish to thank Dr. Florence White of the CCNSC for the data obtained from Dr. Philip S. Thayer of Arthur D. Little, Inc.